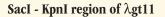
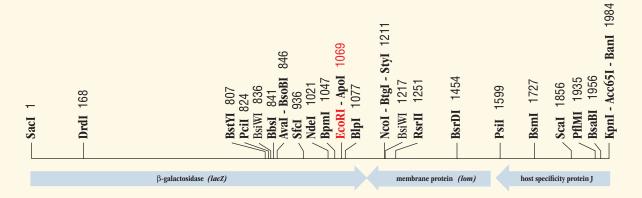
The λ gt11 expression vector was constructed to allow rapid antibody screening of β -galactosidase fusion proteins encoded by DNA cloned into a unique EcoRI site within the 3' end of the *lac2* gene (1,2). Unfortunately, the EcoRI sites are often lost during cloning, making it difficult to excise the insert for subcloning or sequencing. Many researchers have opted to double-digest λ gt11 with SacI and KpnI to liberate a DNA fragment containing inserted DNA at the EcoRI restriction site. The SacI–KpnI fragment can then be subcloned into pUC19 or another suitable vector for sequencing, mapping or further analysis.

NEB has sequenced the SacI-KpnI region of λ gt11 (3). Previously, this sequence was deduced from the sequences of the various constructs used to create the phage. This sequence agrees with the *lacZ* gene sequence and the *lacZ*/ λ fusion junction of $\lambda p / ac 5$ and $\lambda c / 857$ (precursors of $\lambda g t 11$) (4-7). The sequence also contains two open reading frames (ORFs) (the first consists of two thirds of the membrane protein lom fused to a normally untranslated ORF and the second is the host specificity protein J) in the opposite orientation from the *lacZ* ORF, possibly accounting for the occasionally observed translation of proteins in that orientation (5). Several unique restriction targets in the SacI-KpnI sequence have been identified. These may be helpful for restriction mapping DNA cloned into the EcoRI cloning site. In addition, several enzymes have been identified which cleave the SacI-KpnI region twice, once on each side of the EcoRI cloning site. These may be useful for subcloning the region surrounding the EcoRI cloning site into other vectors should the EcoRI site contain multiple inserts or the SacI and KpnI sites be inconvenient (for example if the cloned DNA contains a SacI or KpnI site). The enzyme ApoI (Pu/AATTPy) may be useful for removing some fragments with mutated EcoRI sites.

Positions of sequencing primers available from NEB (not drawn to scale) and positions of restriction endonucleases which cleave the SacI-KpnI region only once are presented in the figure below. The accompanying chart lists those restriction endonucleases which cleave the fragment either once or twice and also provides the recognition site coordinates. Those which cleave twice and whose sites flank the EcoRI site are followed by an asterisk (*). Of particular interest are the locations of the BsiWI sites which cleave only 233 and 148 base pairs to the left and right of the EcoRI position; the BsiWI fragment can then be cloned for further analysis.

Single Sites	Location	Two Sites	Locations	
Acc65I	1984	AccI	828	1706
ApoI	1069	AgeI	1249	1428
AvaI	846	AleI	350	896
BanI	1984	AlwI	807	1758
BbsI*	841	ApaLI	410	525
BlpI	1077	AvaII	1185	1252
BpmI	1047	BaeI	1426	1803
BsaBI	1956	BanII	1	1050
BsmI	1727	BciVI	1303	1432
BsoBI	846	BglI	332	1201
BsrDI	1454	BsaHI	212	384
BstYI	807	BsaXI*	1301	1522
BtgI	1211	BsiWI	836	1217
DrdI	168	BsmFI*	252*	871
EcoRI	1069	BsrBI*	566	730
KpnI	1984	BstXI	278	895
NcoI	1211	BstZ17I	828	1706
NdeI	1021	BtgZI*	967*	1692
PciI	824	BtsI	207	522
PflMI	1935	EaeI	1208	1256
PsiI	1599	HincII	941	1786
RsrII	1251	HinfI	1042	1421
SacI	1	MboII*	842*	988
Scal	1856	MluI	147	572
SfcI	936	MlyI*	1042*	1421
StyI	1211	NlaVI	1049	1984
TspGWI(x)*	1378	NspI	824	1782
(x) = enzyme not available		Nt.BstNBI*	1042*	1421
from NEB		PleI*	1042*	1421
NON NED		PvuII	712	1075
		TatI (x)	1176	1856





 λ gt11 Forward Primer (#S1218S)

 λ gt11 Reverse Primer (#S1222S)

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