

Endonuclease VIII



1-800-632-7799
info@neb.com
www.neb.com



M0299S 009151216121

M0299S



1,000 units **10,000 U/ml** **Lot: 0091512**
RECOMBINANT **Store at -20°C** **Exp: 12/16**

Description: Endonuclease VIII from *E. coli* acts as both an N-glycosylase and an AP-lyase. The N-glycosylase activity releases damaged pyrimidines from double-stranded DNA, generating an apurinic (AP site). The AP-lyase activity cleaves 3' and 5' to the AP site leaving a 5' phosphate and a 3' phosphate. Damaged bases recognized and removed by Endonuclease VIII include urea, 5, 6-dihydroxythymine, thymine glycol, 5-hydroxy-5-methylhydantoin, uracil glycol, 6-hydroxy-5, 6-dihydrothymine and methyltartronylurea (1,2).

While Endonuclease VIII is similar to Endonuclease III, Endonuclease VIII has β and δ lyase activity while Endonuclease III has β lyase activity.

Source: An *E. coli* strain which carries the cloned *nei* gene

Applications:

- Single cell gel electrophoresis (Comet assay) (3,4,5)
- Alkaline elution (6)
- Alkaline unwinding (7)

Supplied in: 250 mM NaCl, 10 mM Tris-HCl (pH 8.0), 0.1 mM EDTA and 50% glycerol.

Reagents Supplied with Enzyme:
10X Endonuclease VIII Reaction Buffer.

Reaction Conditions: 1X Endonuclease VIII Reaction Buffer. Incubate at 37°C.

Endonuclease VIII



1-800-632-7799
info@neb.com
www.neb.com



M0299S 009151216121

M0299S



1,000 units **10,000 U/ml** **Lot: 0091512**
RECOMBINANT **Store at -20°C** **Exp: 12/16**

Description: Endonuclease VIII from *E. coli* acts as both an N-glycosylase and an AP-lyase. The N-glycosylase activity releases damaged pyrimidines from double-stranded DNA, generating an apurinic (AP site). The AP-lyase activity cleaves 3' and 5' to the AP site leaving a 5' phosphate and a 3' phosphate. Damaged bases recognized and removed by Endonuclease VIII include urea, 5, 6-dihydroxythymine, thymine glycol, 5-hydroxy-5-methylhydantoin, uracil glycol, 6-hydroxy-5, 6-dihydrothymine and methyltartronylurea (1,2).

While Endonuclease VIII is similar to Endonuclease III, Endonuclease VIII has β and δ lyase activity while Endonuclease III has β lyase activity.

Source: An *E. coli* strain which carries the cloned *nei* gene

Applications:

- Single cell gel electrophoresis (Comet assay) (3,4,5)
- Alkaline elution (6)
- Alkaline unwinding (7)

Supplied in: 250 mM NaCl, 10 mM Tris-HCl (pH 8.0), 0.1 mM EDTA and 50% glycerol.

Reagents Supplied with Enzyme:
10X Endonuclease VIII Reaction Buffer.

Reaction Conditions: 1X Endonuclease VIII Reaction Buffer. Incubate at 37°C.

1X Endonuclease VIII Reaction Buffer:

10 mM Tris-HCl
75 mM NaCl
1 mM EDTA
pH 8.0 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to cleave 1 pmol of a 34 mer oligonucleotide duplex containing a single AP site* in a total reaction volume of 10 μ l in 1 hour at 37°C.

* An AP site is created by treating 10 pmol of a 34 mer oligonucleotide duplex containing a single uracil residue with 1 unit of Uracil-DNA Glycosylase (UDG) for 2 minutes at 37°C.

Assay Conditions: 1X Endonuclease VIII Reaction Buffer containing 10 pmol of fluorescently labeled oligonucleotide duplex in a total reaction volume of 10 μ l.

Recommended Dilution for Comet Assay:

1:10⁴ to 1:10⁵ (3,4,5,8). A detailed protocol can be found at www.neb.com.

1X Endonuclease VIII Reaction Buffer:

10 mM Tris-HCl
75 mM NaCl
1 mM EDTA
pH 8.0 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to cleave 1 pmol of a 34 mer oligonucleotide duplex containing a single AP site* in a total reaction volume of 10 μ l in 1 hour at 37°C.

* An AP site is created by treating 10 pmol of a 34 mer oligonucleotide duplex containing a single uracil residue with 1 unit of Uracil-DNA Glycosylase (UDG) for 2 minutes at 37°C.

Assay Conditions: 1X Endonuclease VIII Reaction Buffer containing 10 pmol of fluorescently labeled oligonucleotide duplex in a total reaction volume of 10 μ l.

Recommended Dilution for Comet Assay:

1:10⁴ to 1:10⁵ (3,4,5,8). A detailed protocol can be found at www.neb.com.

Quality Control Assays

Physical Purity: Purified to > 95% homogeneity as determined by SDS-PAGE analysis using Coomassie Blue detection.

16-Hour Incubation: A 50 μ l reaction containing 1 μ g of λ DNA (HindIII digest) and 25 units of Endonuclease VIII for 16 hours at 37°C resulted in a DNA pattern free of detectable nuclease degradation as determined by agarose gel electrophoresis.

Exonuclease Activity: Incubation of a 50 μ l reaction containing 10 units of Endonuclease VIII with 1 μ g of a mixture of single and double-stranded [³H] *E. coli* DNA (10⁵ cpm/ μ g) for 4 hours at 37°C released < 0.4% of the total radioactivity

Heat Inactivation: 250 units of enzyme were inactivated by incubation at 75°C for 10 minutes.

Usage Note: Endonuclease VIII will remove deoxyribose- 5' phosphate dR5'P at a nicked site (9).

(see other side)

CERTIFICATE OF ANALYSIS

Quality Control Assays

Physical Purity: Purified to > 95% homogeneity as determined by SDS-PAGE analysis using Coomassie Blue detection.

16-Hour Incubation: A 50 μ l reaction containing 1 μ g of λ DNA (HindIII digest) and 25 units of Endonuclease VIII for 16 hours at 37°C resulted in a DNA pattern free of detectable nuclease degradation as determined by agarose gel electrophoresis.

Exonuclease Activity: Incubation of a 50 μ l reaction containing 10 units of Endonuclease VIII with 1 μ g of a mixture of single and double-stranded [³H] *E. coli* DNA (10⁵ cpm/ μ g) for 4 hours at 37°C released < 0.4% of the total radioactivity

Heat Inactivation: 250 units of enzyme were inactivated by incubation at 75°C for 10 minutes.

Usage Note: Endonuclease VIII will remove deoxyribose- 5' phosphate dR5'P at a nicked site (9).

(see other side)

CERTIFICATE OF ANALYSIS

References:

1. Dizdaroglu, M. Laval, J. and Boiteux, S. (1993) *Biochemistry* 32, 12105–12111.
2. Hatahet, Z. et al. (1994) *J. Biol. Chem.* 269, 18814–18820.
3. Singh, N. et al. (1988) *Experimental Cell Research* 175, 184–191.
4. Collins, A. et al. (1993) *Carcinogenesis* 14, 1733–1735.
5. Collins, A. et al. (1996) *Environmental Health Perspectives* 104, 465–469.
6. Pflaum, M. et al. (1998) *Free Rad. Res.* 29, 585–594.
7. Hartwig, A. et al. (1996) *Toxicology Letters* 88, 85–90.
8. Marks, K., New England Biolabs, Inc., unpublished observations.
9. Marks, K., New England Biolabs, Inc., unpublished observations.



NEW ENGLAND BIOLABS® is a registered trademark of New England Biolabs, Inc.

This product is intended for research purposes only. This product is not intended to be used for therapeutic or diagnostic purposes in humans or animals.

References:

1. Dizdaroglu, M. Laval, J. and Boiteux, S. (1993) *Biochemistry* 32, 12105–12111.
2. Hatahet, Z. et al. (1994) *J. Biol. Chem.* 269, 18814–18820.
3. Singh, N. et al. (1988) *Experimental Cell Research* 175, 184–191.
4. Collins, A. et al. (1993) *Carcinogenesis* 14, 1733–1735.
5. Collins, A. et al. (1996) *Environmental Health Perspectives* 104, 465–469.
6. Pflaum, M. et al. (1998) *Free Rad. Res.* 29, 585–594.
7. Hartwig, A. et al. (1996) *Toxicology Letters* 88, 85–90.
8. Marks, K., New England Biolabs, Inc., unpublished observations.
9. Marks, K., New England Biolabs, Inc., unpublished observations.



NEW ENGLAND BIOLABS® is a registered trademark of New England Biolabs, Inc.

This product is intended for research purposes only. This product is not intended to be used for therapeutic or diagnostic purposes in humans or animals.