

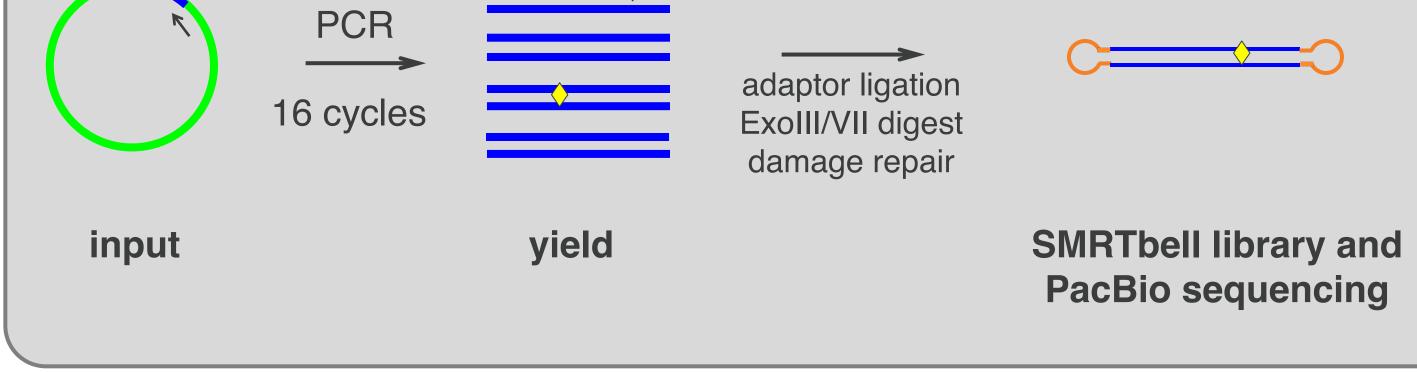
Examining Sources of Error in PCR by Single-Molecule Sequencing

Jennifer L. Ong and Vladimir Potapov New England Biolabs, Ipswich, MA 01938, USA

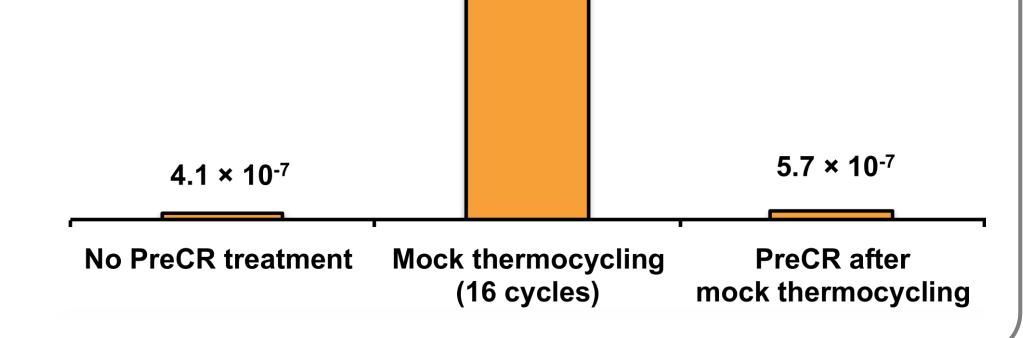
INTRODUCTION

The Polymerase Chain Reaction (PCR) is an integral part of many NGS sample preparation workflows. Mistakes made during PCR appear in sequencing data and contribute to false mutations that can ultimately confound genetic analysis. We utilized a single-molecule sequencing assay to comprehensively catalog the different types of errors introduced during PCR, including polymerase misincorporation, structure-induced template-switching, PCR-mediated recombination and DNA damage caused by thermocycling.

| METHODS | RESULTS | | | |
|-----------------------|--|------------------------|--|--|
| EXPERIMENTAL STRATEGY | SEQUENCEABLE DNA DAMAGE FROM THERMOCYCLING | 2.3 × 10 ⁻⁵ | | |
| | Plasmid libraries were subjected to 16 mock thermocycles and sequenced before and after thermocycling. | | | |



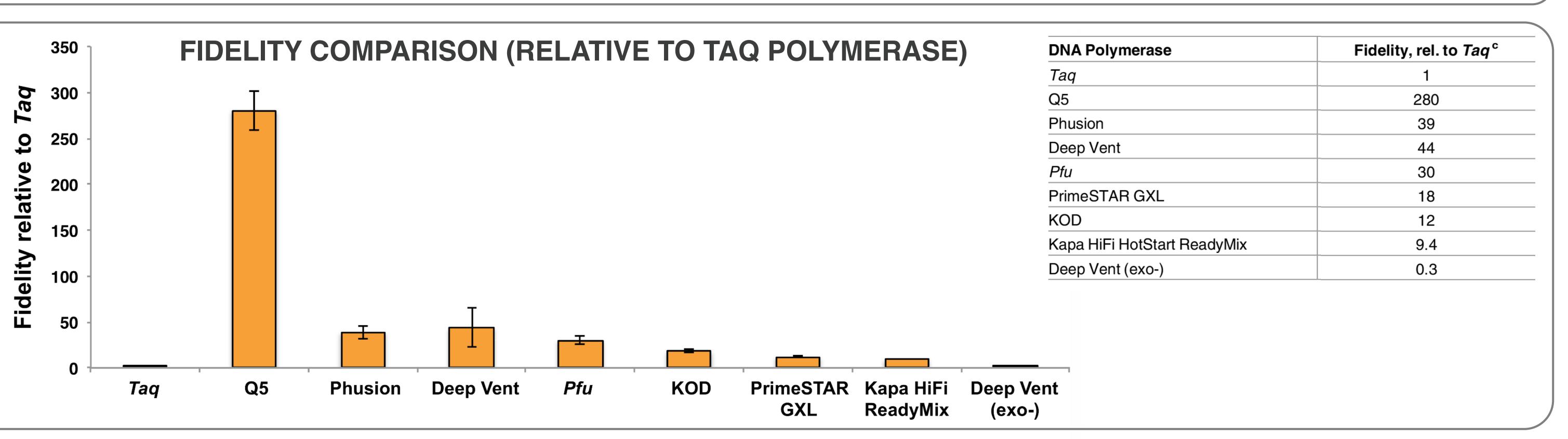
- and alter thermosyomig,
- Per cycle, thermocycling introduced mutations at a rate of once per 714,000 bases, almost all mutations (97%) were C->T, and indicative of cytosine deamination
- PreCR treatment (DNA damage repair) after mock thermocycling reduces substitution rate to pre-thermocylcing levels



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BIOINFORMATICS WORKFLOW

- For each ZMW, split all subreads into two groups (top or bottom strand) based on mapping directionality (BLASR) and build consensus reads (CCS 2)
- Map consensus reads (BLASR) and tally up all mutations by comparing consensus read to a reference
- Read filtering: number of passes >15 and Quality Value =93; length of consensus read >80% of the reference; exclude chimeric reads (BWA)
- Compute error rates and normalize to number of doubling events in PCR



POLYMERASE BASE SUBSTITUTION ERROR RATES

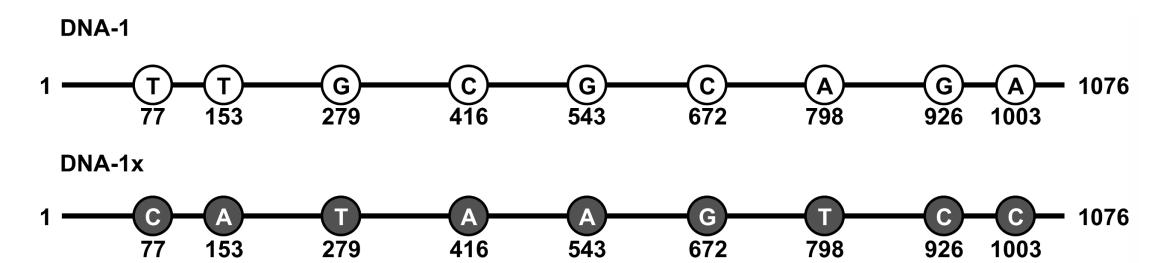
| DNA Polymerase | Substitution rate ^a | Accuracy ^b |
|-----------------------------|--|-----------------------|
| Taq | $1.5 \times 10^{-4} (\pm 0.2 \times 10^{-4})$ | 6,456 |
| Q5 | $5.3 \times 10^{-7} (\pm 0.9 \times 10^{-7})$ | 1,870,763 |
| Phusion | $3.9 \times 10^{-6} (\pm 0.7 \times 10^{-6})$ | 255,118 |
| Deep Vent | $4.0 \times 10^{-6} (\pm 2.0 \times 10^{-6})$ | 251,129 |
| Pfu | 5.1 × 10 ⁻⁶ (± 1.1 × 10 ⁻⁶) | 195,275 |
| PrimeSTAR GXL | 8.4 × 10 ⁻⁶ (± 1.1 × 10 ⁻⁶) | 118,467 |
| KOD | $1.2 \times 10^{-5} (\pm 0.2 \times 10^{-5})$ | 82,303 |
| Kapa HiFi HotStart ReadyMix | $1.6 \times 10^{-5} (\pm 0.3 \times 10^{-5})$ | 63,323 |
| Deep Vent (exo-) | $5.0 \times 10^{-4} (\pm 0.1 \times 10^{-4})$ | 2,020 |

MUTATIONAL SPECTRUM FOR VARIOUS POLYMERASES 20% 0% 40% 60% 80% 100% Taq 68 **Q5** Phusion Deep Vent 50 23 Pfu PrimeSTAR GXL KOD 45 Kapa HiFi ReadyMix 76 Deep Vent (exo-)

 $\square A \rightarrow G, T \rightarrow C \square G \rightarrow A, C \rightarrow T \square A \rightarrow T, T \rightarrow A \square A \rightarrow C, T \rightarrow G \square G \rightarrow C, C \rightarrow G \square G \rightarrow T, C \rightarrow A$

^a Substitution rate: substitutions/base/doubling

^b Accuracy: number of bases over which 1 substitution error is expected



TEMPLATE-SWITCHING BY TAQ DNA POLYMERASE

A single-molecule assay to measure template-switching during amplification of a mixed population by Taq polymerase

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Strands with at least 1 recombination event

| | Template pair | N _{re} ^a | N _{total} ^D | Recombination rate ^c | Strands with at least 1 recombination event | | | |
|--|--|------------------------------|---------------------------------|---------------------------------|---|--|--|--|
| PCR amplification | DNA-1:DNA-1x | 19,943 | 77,725,936 | 9.6 × 10 ⁻⁵ | 23% | | | |
| No recombination | DNA-2:DNA-2x | 14,687 | 44,271,304 | 1.3 × 10 ⁻⁴ | 28% | | | |
| 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - | ^a Number of recombination events. ^b Total number of analyzed sequenced bases. ^c Recombination rate is per base per doubling. Recombination rate is doubled to account for "cryptic" recombination events. | | | | | | | |
| REFERENCES | CONCLUSIONS | | | | | | | |
| | | | | | | | | |
| Potapov V, Ong JL (2017) Examining Sources of Error in PCR by Single- Molecule Sequencing. PLOS ONE 12(1): e0169774. doi: 10.1371/journal.pone.0169774 | We analyzed PCR products at the single-molecule level and present here a more complete picture of the types of mistakes that occur during DNA amplification. In addition to polymerase base substitution errors, other sources of error were found to be equally prevalent. For very accurate polymerases, DNA damage introduced during temperature cycling appear to be a major contributor towards mutations occurring in amplification products. PCR-mediated recombination by Taq polymerase was observed at the single-molecule level, and found to occur as frequently as polymerase base substitution errors, suggesting it may also be an underappreciated source of error for multiplex amplification reactions | | | | | | | |
| https://github.com/potapovneb/pcr-fidelity | | | | | | | | |